

CoA written by:	Mahfuz Chowdhury	CoA approved by:	Andrew Prowse
Signature:	<i>Mahfuz</i>	Signature:	<i>A Prowse</i>
Date:	01 Aug 2024	Date:	01Aug2024

## Product Information and Certificate of Analysis

### Human Embryonic Stem Cell Line MEL-1

#### Product Description

Originating from Australia, the MEL-1 cell line was derived by Stem Cell Sciences Ltd. in a collaboration with Melbourne IVF in an NHMRC-licensed project from 'excess' human embryos donated by couples having IVF treatment. This line is registered on the USA's National Institute of Health's [Human Embryonic Stem Cell Registry](#).

#### Properties

Cell line	MEL-1
Cell type	Human embryonic stem cells
Gender	Male
Bank designation	Working Cell Bank (WCB) #4
Storage conditions	≤ -135°C
Date of cryopreservation	22/05/2024
Passage number	P18
Cell number	4 × 10 <sup>6</sup> cells per vial
Cryopreservation medium	CryoStor® CS10
Culture medium	mTeSR™ Plus
Matrix substrate	Matrigel® - hESC-Qualified Matrix
Recommended passage method and split ratio	ReLeSR™, cells can generally be split 1:8-1:12 every 4 days

#### Recommended Materials for Use

Reagent	Supplier	Cat. No.
ReLeSR™	STEMCELL Technologies	100-0484
mTeSR™ Plus Kit	STEMCELL Technologies	05825
RevitaCell Supplement (100X)	Thermo Fisher Scientific	A2644501
Corning® Matrigel® hESC-Qualified Matrix	Corning®	354277
Accutase™ Cell Dissociation Reagent	Thermo Fisher Scientific	A1110501

## Certificate of Analysis

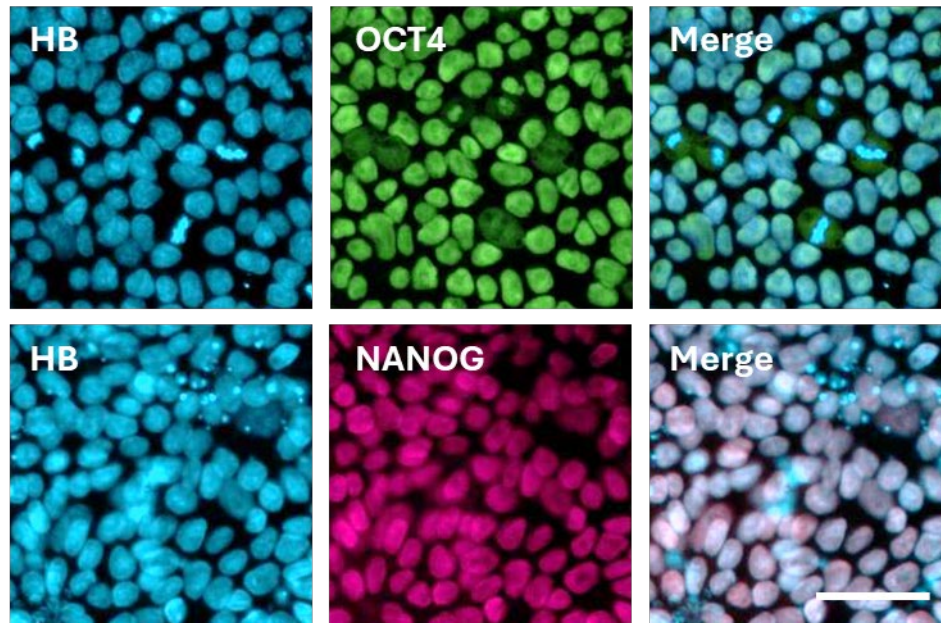
The following testing specifications have been met for this product:

Test Description	Test Method	Test Results
<b>Post-thaw cell viability and recovery</b>	Cell count	Viable cell count at revival > 80%. Cells reached > 80% confluency within 4 days when one stock vial cultured on a Matrigel-coated T75 flask.
<b>Mycoplasma</b>	Bioluminescence (Lonza MycoAlert detection kit)	Negative
<b>Expression of pluripotent stem cell markers</b>	Immunofluorescence	Positive for OCT4 and NANOG
	Flow cytometry	> 90% positive (SSEA4 and TRA-1-60)
	Undifferentiated state marker expression	Comparable expression of SOX2, NANOG, OCT4, DNMT3B and REX1 to control
<b>Genetic Analysis</b>	Copy number variations (CNV) test	No CNVs ( $\leq$ 1 Mb gain/loss tested in 9 loci) detected
	Karyostat assay	No CNVs (1-2 Mb gain or loss tested in 18,018 loci) detected.
<b>Differentiation potential</b>	Immunofluorescence	<u>Positive for lineage specific markers:</u> Ectoderm (NESTIN, PAX6) Mesoderm (NCAM1, BRACHYURY) Endoderm (FOXA2, SOX17)
	Gene expression by RT-qPCR	<u>Increased expression of lineage-specific markers:</u> Ectoderm (PAX6, SOX1, NR2F2) Mesoderm (HAND1, T, CDX2) Endoderm (SOX17, FOXA2, GATA6)

## 1. Expression of Pluripotent Stem Cell Markers

### 1.1. Immunofluorescence

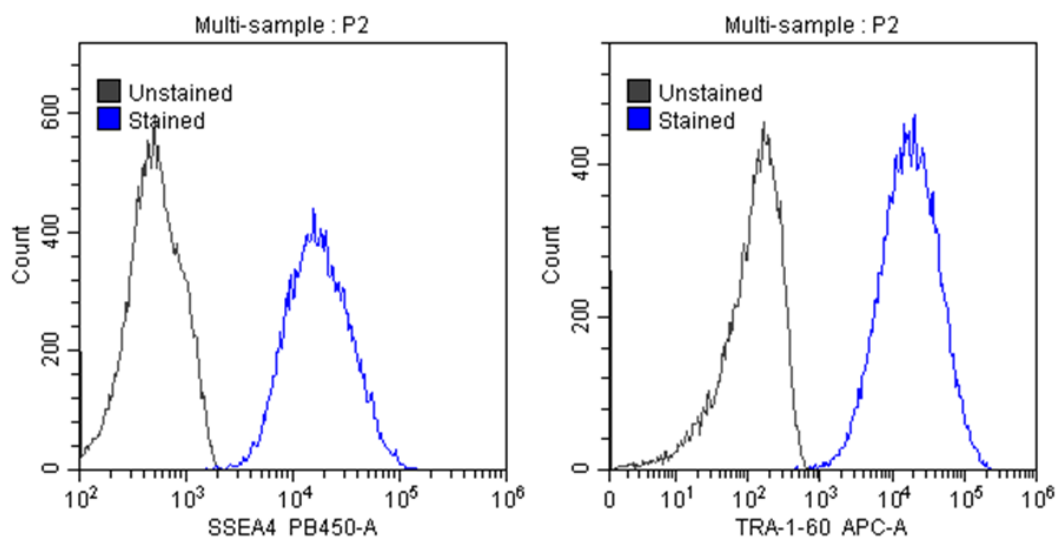
To assess undifferentiated state, MEL-1 cells were stained for nuclear markers OCT4 and NANOG.



**Figure 1.** Expression of marker for the undifferentiated state, OCT4 and NANOG in MEL-1 hES cells detected by immunostaining. HB: Hoechst Blue; Scale bar represents 50  $\mu$ m.

### 1.2. Flow Cytometry

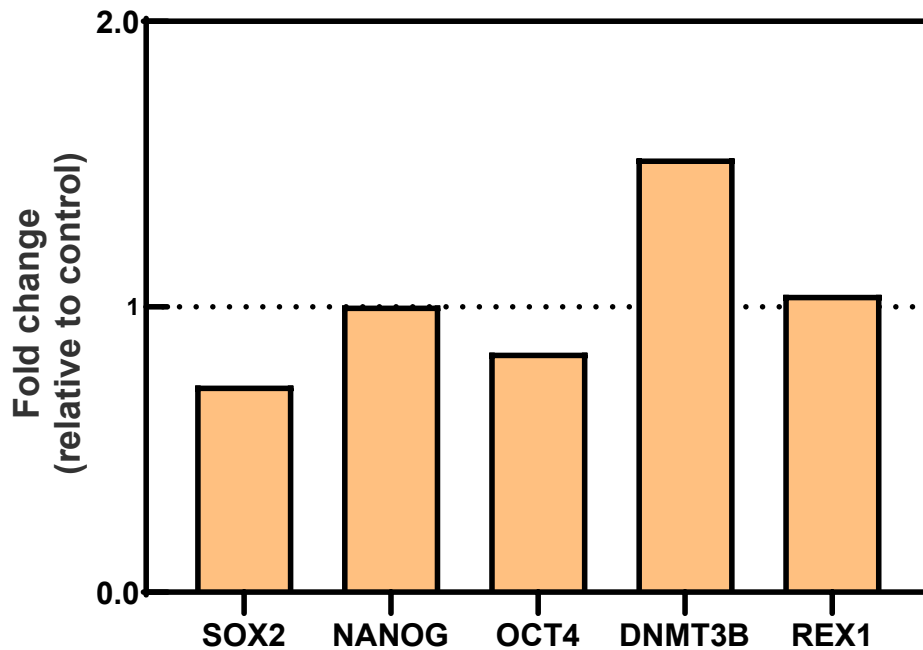
In addition to immunofluorescence, flow cytometry analysis confirmed that > 90% of cells expressed undifferentiated state markers SSEA4 and TRA-1-60.



**Figure 2.** Expression of undifferentiated cell markers SSEA4 (left box) and TRA-1-60 (right box) in the cell line detected by flow cytometry. Black and blue histograms represent unstained and stained cells, respectively.

### 1.3. Gene Expression Analysis

Relative gene expression was measured by Reverse Transcription Quantitative PCR (RT-qPCR) using mRNA extracted from the cells. GAPDH was used as the endogenous housekeeping gene. Relative gene expression demonstrated that pluripotent gene expression was comparable between MEL-1 and MEL-3 (used as the comparison control) hES cells.

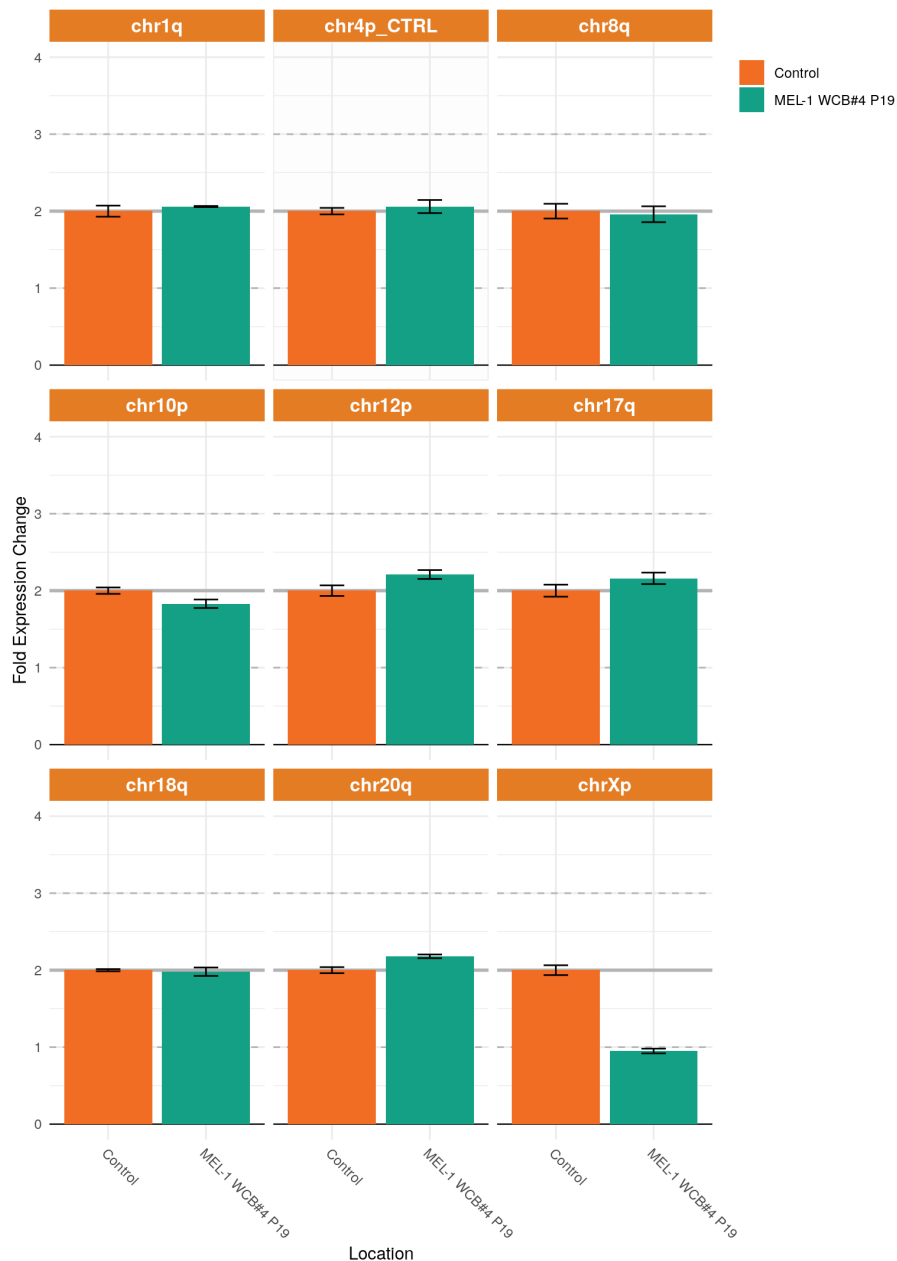


**Figure 3.** Gene expression analysis of pluripotency markers in MEL-1 cells demonstrating the expression of SOX2, NANOG, OCT4, DNMT3B and REX1. Expression (dotted line) in MEL-3 hES line represents positive pluripotent control. Data represents mean for three technical replicates from one biological replicate.

## 2. Genetic Analysis

### 2.1 Copy Number Variation (CNV) Test

MEL-1 hES cells were screened for recurrent CNVs commonly reported in hES cells using hPSC Genetic Analysis Kit (STEMCELL Technologies). Genomic DNA was extracted, and qPCR was performed using the kit. qPCR data was analysed using the appicate available at [STEMCELL Technologies Genetic Analysis Tool](#). MEL-1 WCB#4 cells showed no abnormalities ( $\leq 1$  Mb gains) in all tested loci. Presence of one copy of chrXp confirmed the male sex identity of MEL-1 cells.

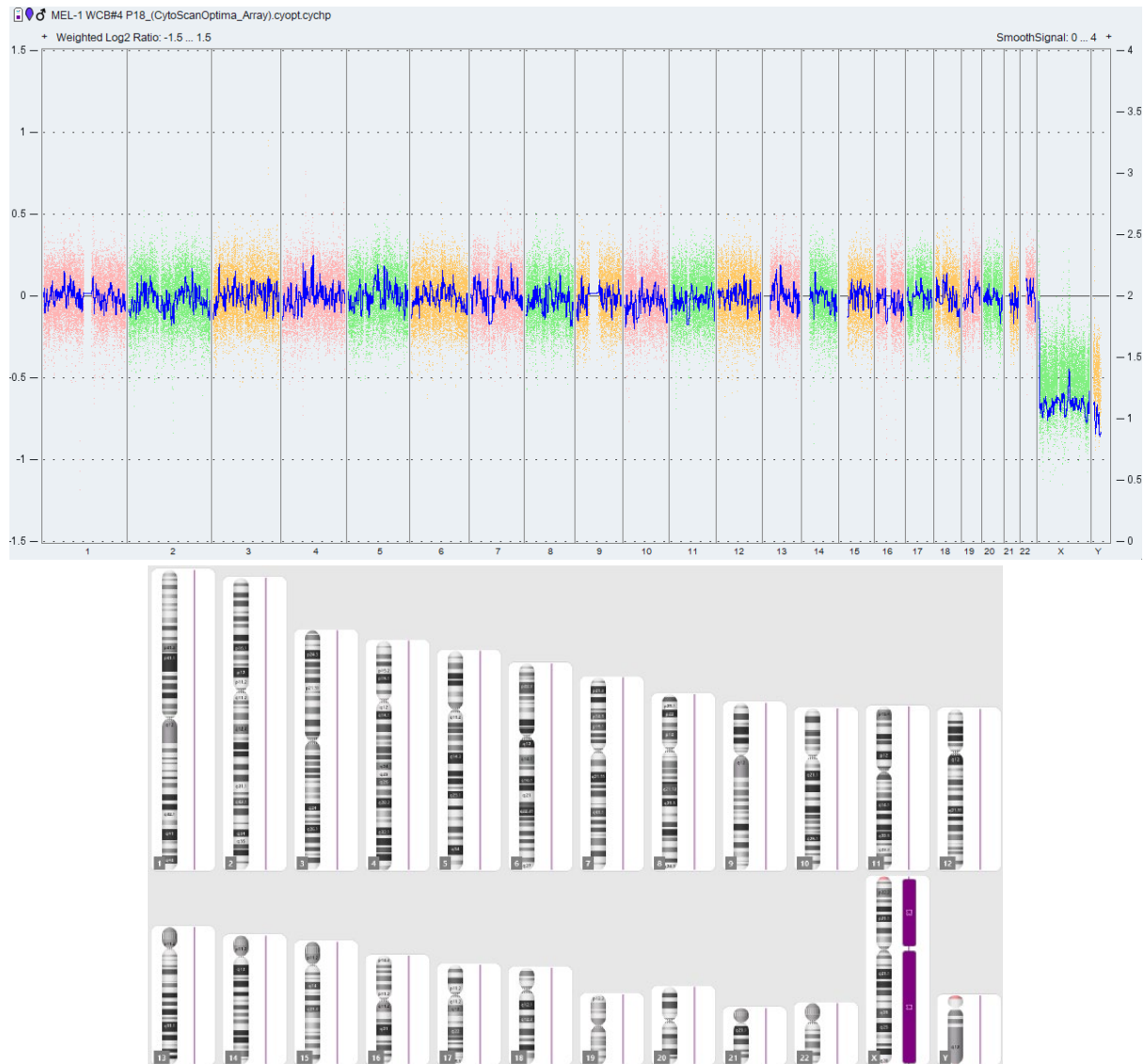


**Figure 4.** Copy Number Variation test result summary for MEL-1 hESCs WCB#4. Data represents mean  $\pm$  SD for three technical replicates of each sample.

## 2.2 KaryoStat Assay

The KaryoStat Assay provides much better resolution than G-banding karyotyping offering detection of multiple types of chromosomal abnormalities in the whole-genome. The KaryoStat assay on MEL-1 WCB#4 sample was performed by using [CytoScan Optima Suite](#) that can detect copy number variations (CNVs) in 18,018 regions, single-nucleotide polymorphism (SNP) in 148,450 regions as well as allelic imbalance, loss of heterozygosity (LOH).

The analysis of the assay data demonstrated that there was no detectable CNVs (1-2 Mb gain or loss) in any chromosomes.



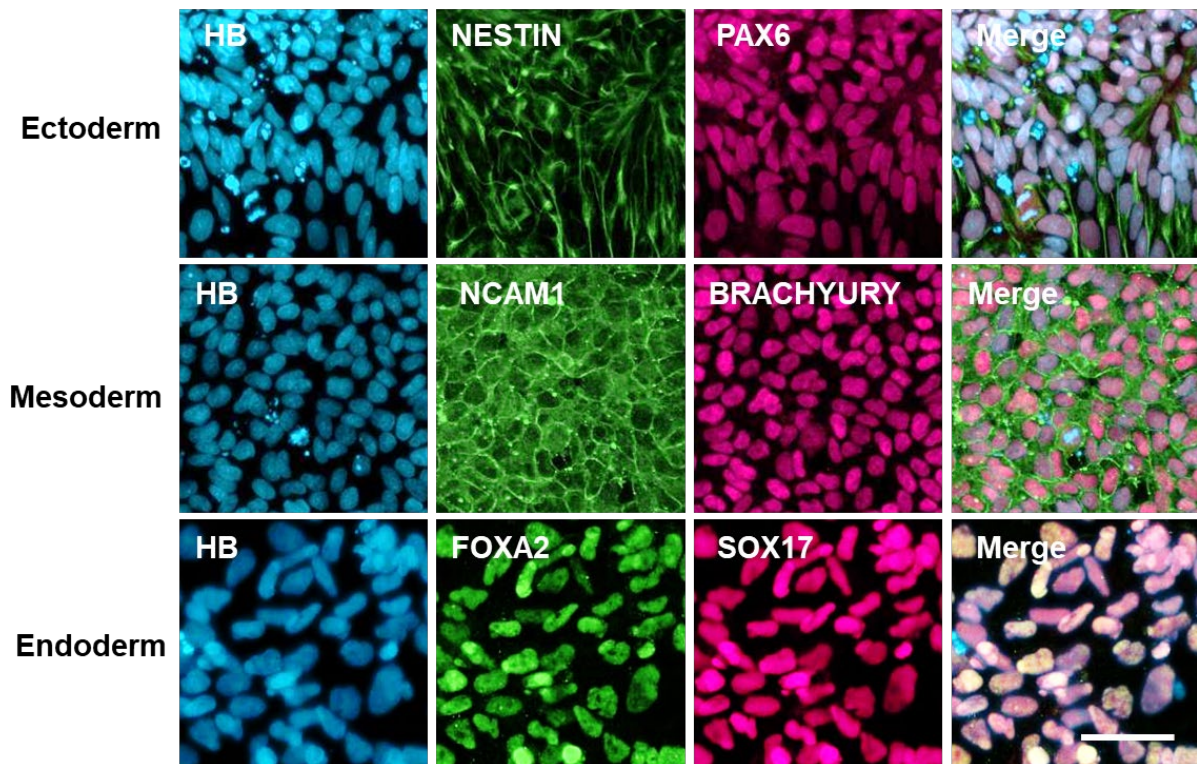
**Figure 5.** Summary of the KaryoStat assay data analysed with ChAS 4.4. Top panel shows fold change (right y-axis, blue line indicates mean of the signal) of the regions analysed for CNVs. Mean value hovering around value of 2 or 1 (for X and Y chromosomes, because of male identity of the cells) indicates absence of CNVs in MEL-1 WCB#4 cells. Karyoview (bottom panel) also confirm the absence of any loss or gain.

### 3. Differentiation Potential Analysis

Pluripotent state of MEL-1 cells was confirmed by differentiating them in three germ layers (Ecto, Meso and Endoderm) by using the STEMdiff Trilineage Differentiation Kit (STEMCELL Technologies) and characterising the differentiated cells with immunofluorescence and qPCR.

#### 3.1. Immunofluorescence

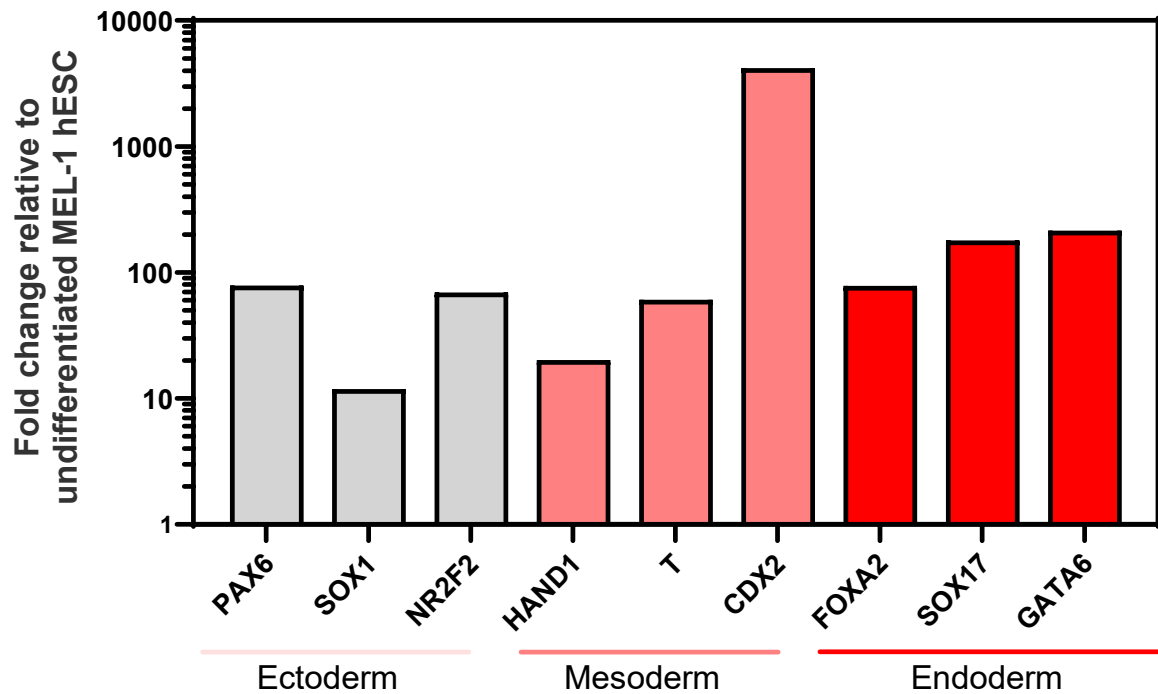
Differentiated cells were stained for two markers of each germ layer.



**Figure 6.** Trilineage differentiation of MEL-1 cells demonstrating the presence of markers from the three germ layers – Ectoderm (NESTIN and PAX6), Mesoderm (NCAM1 and BRACHYURY) and Endoderm (FOXA2 and SOX17) and. HB: Hoechst blue; Scale bar represents 50  $\mu$ m.

### 3.2. Gene Expression Analysis

Relative gene expression was measured from extracted mRNA by using RT-qPCR. GAPDH was used as the endogenous housekeeping gene.



**Figure 7.** Gene expression analysis of differentiated MEL-1 cells demonstrating the expression of genes from the three germ layers - Ectoderm (PAX6, SOX1 and NR2F2), Mesoderm (HAND1, T and CDX2) and Endoderm (FOXA2, SOX17 and GATA6) relative to undifferentiated control (expression level = 1). Data represents mean for three technical replicates of each sample.

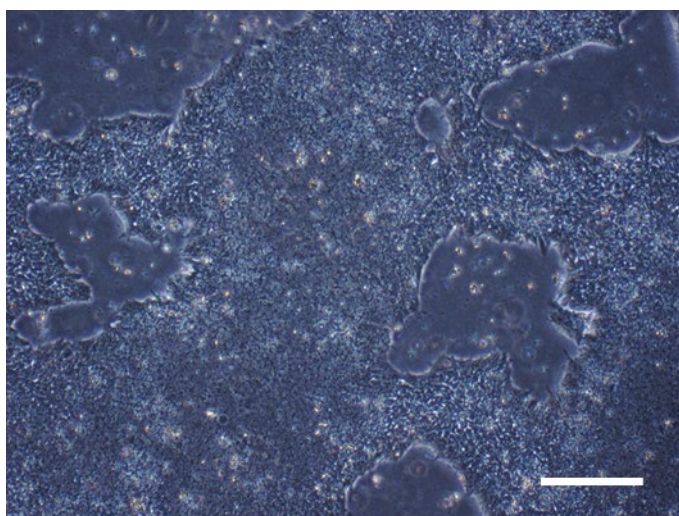


## 4. Directions for Use

The following instructions are for preparing one cryovial of MEL-1 cells for plating in  $1 \times 75\text{cm}^2$  flask. Use aseptic technique throughout the protocol.

### 4.1. Thawing and plating MEL-1 hES cells

- 1) Coat culture-ware flask with Matrigel® (made to manufacturer's specifications) and incubate for at least 30 min at  $37^\circ\text{C}$  or overnight at  $4^\circ\text{C}$ . Prior to cell seeding, the flask should be brought to room temperature.
- 2) Aliquot required volume of mTeSR plus media (~ 15 mL – 20 mL for seeding in one T75) in a 50 mL tube, add Y-27632 (i.e., rock inhibitor) for 10  $\mu\text{M}$  final concentration and pre-warm to  $37^\circ\text{C}$ . Use this media in steps 3 to 9.
- 3) Gently warm the cryovial in a water bath until a small ice pellet remains and transfer into a biosafety cabinet.
- 4) Add 1 mL of mTeSR media to cryovial and transfer content into a 15 mL centrifuge tube containing 2 mL of mTeSR media.
- 5) Centrifuge at  $300 \times g$  for 3 min.
- 6) Aspirate the media avoiding disruption to the cell pellet and add 1 mL of mTeSR media.
- 7) Remove the substrate coating prior to transferring the cells and add mTeSR to flask (11 mL/T75 flask).
- 8) Gently re-suspend the cell pellet and transfer to the flask.
- 9) Transfer flask into an incubator at  $37^\circ\text{C}$  and do not disturb it for 24 hrs.
- 10) Perform a full media exchange after 24 hrs (without Y-27632) and then every other day.



**Figure 8.** MEL-1 hESC colonies at Day 4 post-recovery from cryostorage. Scale bar 200  $\mu\text{m}$ .

## 4.2. Passaging MEL-1 hES cells

- 1) When cells reach 80-90% confluence prepare the following reagents and volumes per T75 flask (adjust volumes as required for specific culture flask):
- 2) 5 mL ReLeSR™ (pre-warm to 37°C).
- 3) 10 mL of PBS (calcium and magnesium free (CMF), pre-warm to 37°C).
- 4) T75 flasks pre-coated with extracellular matrix (ECM) of choice according to manufacturer's instructions.
- 5) 18 mL mTeSR™ Plus (pre-warm to 37°C) containing 10 µM Y-27632.
- 6) 10 mL of DMEM (pre-warm to 37°C, specific formulation not strictly important).
- 7) Remove flask from incubator and transfer to biosafety cabinet.
- 8) Aspirate media and wash cells with 10 mL PBS CMF. Aspirate PBS.
- 9) Add 5 mL of ReLeSR™. Lay down flask and ensure entire surface is covered by ReLeSR™.
- 10) Incubate in biosafety cabinet for no more than 30 sec.
- 11) Aspirate ReLeSR™ and immediately transfer flask to 37°C incubator.
- 12) Incubate for 2 min. Check cells under microscope for rounding of cells. Incubate for 1 min longer if required.
- 13) In the biosafety cabinet, add 5 mL of DMEM down the side of the flask.
- 14) Tilt flask so surface is covered with DMEM and tap the side of the flask 2 or 3 times. If incubation time in ReLeSR™ is sufficient, cells will lift off with tapping. Collect the cell suspension in a 15 mL centrifuge tube.
- 15) Wash the surface with another 5 mL DMEM and transfer to the centrifuge tube.
- 16) Pellet the cells at  $300 \times g$  for 3 min.
- 17) Aspirate supernatant and add 5 mL of mTeSR™ Plus with Y-27632. Mix well to resuspend pellet.
- 18) Aspirate ECM from T75 and immediately add 12 mL of mTeSR™ Plus with Y-27632 down the side of the flask to avoid removing ECM from surface.
- 19) Take 1:8 to 1:12 (or  $0.75 \times 10^6$  to  $1 \times 10^6$  cells) of the cell suspension and transfer to one T75 with media.
- 20) Transfer to CO<sub>2</sub> incubator. Gently agitate flask back and forth to ensure even spread of cells in the flask.
- 21) Exchange media after 24 hrs with 20 mL mTeSR plus without Y-27632 and then every other day.
- 22) Cells will reach 80-90% confluence within 4 days.

### 4.3. Cryopreservation MEL-1 hES cells

- 1) Follow the passaging protocol up to step 17 and take 0.5 or 1 mL of the cell suspension into a 15 mL tube for cell counting.
- 2) Centrifuge the 15 mL tube at  $300 \times g$  for 3 min.
- 3) Resuspend cell count aliquot in 0.5 mL of pre-warmed Accutase™ (include a PBS CMF wash before this if desired. Cells are currently in small clusters of 5 - 10 cells and don't require much effort for single cell dissociation).
- 4) Incubate at  $37^{\circ}\text{C}$  for 3 min with regular gentle agitation. After incubation, add 0.5 mL mTeSR+ media and pipette 5-10 times using a 200  $\mu\text{L}$  pipette.
- 5) Perform cell count using desired method.
- 6) Back calculate to determine cell number of original cell suspension. Depending on confluence, each T75 flask may contain  $15 - 25 \times 10^6$  cells.
- 7) Centrifuge the original cell suspension at  $300 \times g$  for 3 min and aspirate media.
- 8) Gently flick tube to dislodge cell pellet.
- 9) Add cold, CryoStor® CS10 to a final cell density of  $2 - 5 \times 10^6$  cells/mL. We don't recommend cryopreserving densities lower than  $2 \times 10^6$  cells/mL. Higher densities improve cell recovery at thaw.
- 10) Aliquot  $\geq 500 \mu\text{L}$  of cell suspension in CS10 per cryovial.
- 11) Transfer the cryovials to Mr. Frosty Cryo freezing containers, store the containers at  $-80^{\circ}\text{C}$  for overnight and then transfer the vials to LN2 cryostorage.